Actinobacteriophage Genome Annotation Submission Cover Sheet

This Cover Sheet will accompany each genome’s annotation file(s) submission and succinctly describe the work that your students and you have done. This document ensures that the work done was as complete and thorough as it could be. Most important to the QC reviewer, denote where the trouble spots were in your annotation and how they were resolved.

Phage Name. Phegasus

Your Name. Susanne Pfeifer

Your Institution. Arizona State University

Your email. susanne.pfeifer@asu.edu

Additional emails. (for correspondence).

Describe any issues or specific genes that you would like to highlight for the QC reviewer. This includes any genes that you had questions about or received help with or that warrant further inspection in the QC review process. Include those genes that you deliberated on and/or want to strongly advocate for. If you contacted SMART, workshop facilitator, or a buddy school for help, please document.

1. Three new genes (44, 56, 64) not present in the original auto-annotation are supported by BLAST and synteny results, but the supporting coding potential for gene 44 is primarily in the +2 frame rather than the +3 frame where the ORF is, the supporting coding potential for gene 56 is primarily in the -2 frame rather than the -1 frame where the ORF is, and the supporting coding potential for gene 64 is in the correct +3 frame where the ORF is but is weak (<0.5).
2. Genes 9, 17, 22, 25, 37, 41, 45, 50, and 58 all contain tandem start codons where the first start codon produces a better or equivalent BLAST alignment. We selected the second of the two potential start codons as the Bioinformatics guide mentioned that wet bench experiments often support the second of the two codons as the correct start in cases like these.
3. A potential tRNA was found by tRNAscan-SE (Infernal Score 12.6), however, it was within a region with high coding potential ([26972,27076]; within gene 31); thus, we did not call it.

Please record yes/no for each of the questions below. If further explanation is needed, please add this item to the above box.

In the submitted DNA Master file (Yes/No):

1. Does the genome sequence in your submitted DNA Master file match the nucleotide fasta file posted on phagesDB (same number of bases, no N bases, etc.)? **Yes**

2. Are all the genes ‘Valid” when you click the [Validation button](https://seaphagesbioinformatics.helpdocsonline.com/article-84)? **No**

 Frameshift evidence for genes 15-16

 9932 - 10850

 Not In Frame

 1 In-frame Stop Codon

 9932 - 10531 and 9932 - 10850 share a 5' (upstream) coordinate

3. Are the genes (and matching LocusTag numbers) [sequential](https://seaphagesbioinformatics.helpdocsonline.com/article-77), starting with #1, counting by 1s.
**Yes**

4. Are the Locus Tags the “[SEA\_PHAGE NAME](https://seaphagesbioinformatics.helpdocsonline.com/article-77)” format? **Yes**

5. Has the [documentation been recreated](https://seaphagesbioinformatics.helpdocsonline.com/article-86) from the Feature Table to match the latest file version? **Yes**

6. Have tRNAs followed the [tRNA protocol](https://seaphagesbioinformatics.helpdocsonline.com/undefined), **COPYING** tRNA-AMINOACID type (DNA equivalent of the anti-codon) from Aragorn output - ﻿tRNA-Gln(ctg) - AND the ends been adjusted to match the Aragorn output? **N/A**

7. Has the [frameshift in the tail assembly chaperone](https://seaphagesbioinformatics.helpdocsonline.com/article-54) been annotated correctly (if applicable)? **Yes**

8. Have you cleared your Draft\_Blast data and have you [re-Blasted](https://seaphagesbioinformatics.helpdocsonline.com/article-57) the submitted DNA Master file? **Yes**

9. Has every gene been [described and supported in your Supporting Data file](https://seaphagesbioinformatics.helpdocsonline.com/article-44)? **Yes**

10. Did you investigate ‘[gaps](https://seaphagesbioinformatics.helpdocsonline.com/article-31)’? **Yes**

11. Did you [delete the genes](https://seaphagesbioinformatics.helpdocsonline.com/article-65) that you meant to delete? **N/A**

Now, [make a profile of the file](https://seaphagesbioinformatics.helpdocsonline.com/article-64) you plan to send. (And you can save this file for [Review to Improve!)](https://seaphagesbioinformatics.helpdocsonline.com/untitled-18)

1. Have any duplicate genes been deleted? **N/A**

2. Has the Notes field been cleared (using the automated buttons)? **Yes**

3. Do the gene numbers and locus tags match? **Yes**

4. Are the correct Feature\_Types correctly selected (most will be ORFs, but check that tRNAs and tmRNAs are correctly labeled)? **Yes**

5. Do the function names in the Product field either match the official function list or say “Hypothetical Protein”? **Yes**

6. Has the Function field been cleared (using the automated buttons)? **Yes**

How are you documenting your gene calls in class? Choose any/all that apply:

      PECAAN output

      DNA Master shorthand (previously used format)

**X** Spreadsheet

      Powerpoint

      Word document (must be easily searchable)

      Other: Describe.

What is the file type (sort) submitted for QC to document your gene calls? Choose only one.:

      PECAAN output

      DNA Master shorthand (previously used format)

**X** Spreadsheet

      Powerpoint

      Word document (must be easily searchable)

      Other: Describe.